Supplementary Material

Synthesis of a Rhodanine-Based Compound Library Targeting Bcl-XL and Mcl-1

Paul H. Bernardo, Thirunavukkarasu Sivaraman,Kah-Fei Wan, Jin Xu, Janarthanan Krishnamoorthy, Chun Meng Song, Liming Tian, Jasmine S. F. Chin, Diane S. W. Lim, Henry Y. K. Mok, Victor C. Yu, Joo Chuan Tong, and Christina L. L. Chai

Experimental Section

Chemistry. General Experimental Section

All ¹H NMR and ¹³C NMR spectra were acquired on a Bruker 400 UltraShield Spectrometer operating at 400 MHz and 100 MHz respectively. The proton spectra were referenced to the respective residual solvent peaks (MeOH- d_4 : 3.20; DMSO- d_6 : 2.50 ppm) except for those recorded in CDCl₃ which were referenced to TMS. Carbon spectra were referenced to the central peak of the respective residual solvents (MeOH- d_4 : 49.0; DMSO- d_6 : 39.5; CDCl₃: 77.0 ppm). Low-resolution and high-resolution electron impact mass spectra (EIMS) were measured using a Finnigan MAT95XP double-focusing mass spectrometer. Low-resolution electrospray ionization (ESI) mass spectra were recorded using Waters Quattro MicroTM API. High resolution ESI spectra were obtained using the Agilent 6210 Time-of-Flight LC/MS. Infrared (IR) spectra were measured on a BioRad FTIR spectrophotometer with samples analyzed as KBr discs, or as thin films on a KBr plate. All elemental analyses were carried out on a EuroEA3000 series CHNS Analyzer.

Experimental Details. General Procedure for the Synthesis of N-substituted Rhodanines (7a-f)

To a solution of the chosen amino acid (typically 5 g, 30 mmol) in water (100 mL) was added NaOH (2.4 g, 2 equiv.) and CS_2 (1.81 mL, 30 mmol). The reaction mixture was stirred for 16 h at room temperature after which a solution of sodium chloroacetate (2.82 g, 30 mmol) was added. The reaction mixture was stirred for a further 3 h at room temperature, then acidified with aqueous HCl (6 N, 30 mL). The resulting solution was refluxed for 16 h, then cooled to room temperature. The crude product was extracted from the aqueous layer with ethyl acetate (3 x 50 mL), and the combined organic layer was dried with MgSO₄, filtered, and the solvent removed in vacuo. The products were recrystallized from ethanol-water mixtures.

2-(4-oxo-2-thioxothiazolidin-3-yl)acetic acid (**7a**). Yellow solid, 89% yield. Mp 148 °C (lit.[1]148 °C). ¹H NMR (CDCl₃): δ 4.16 (s, 2H, NCH₂), 4.79 (s, 2H, SCH₂).

(*S*)-2-(*4-oxo-2-thioxothiazolidin-3-yl*)*propanoic acid* (**7b**). Yellow solid, 90% yield. Mp 152-153 °C. (lit.[2]152-153 °C). ¹H NMR (CDCl₃): δ 1.61 (d, *J* = 7 Hz, 3H, CH₃), 4.00 (d, *J* = 2 Hz, 2H, CH₂), 5.68 (q, *J* = 7 Hz, 1H, NCH).

(*S*)-3-methyl-2-(4-oxo-2-thioxothiazolidin-3-yl)butanoic acid (**7c**). Yellow solid, 92% yield. Mp 113-115 °C. (lit.[3]113-115 °C). ¹H NMR (CDCl₃): δ 0.81 (d, *J* = 7 Hz, 3H, CH₃), 1.24 (d, *J* = 7 Hz, 3H, CH₃), 2.80 (m, 1H, CH₃CH), 4.03 (s, 2H, SCH₂), 5.25 (d, *J* = 9 Hz, 1H NCH), 9.60 (s, 1H, CO₂H).

(*S*)-4-methyl-2-(4-oxo-2-thioxothiazolidin-3-yl)pentanoic acid (**7d**). Pale yellow solid, 88% yield. Mp 100 °C. (lit.[3] 99-101 °C). ¹H NMR (CDCl₃): δ 0.91 (d, *J* = 7 Hz, 3H, CH₃), 0.95 (d, *J* = 7 Hz, 3H, CH₃), 1.51 (m, 1H, CH₃CH), 2.09 (m, 2H, NCHCH₂), 3.98 (s, 2H, SCH₂), 5.66 (m, 1H, NCH).

(2*S*,3*S*)-3-methyl-2-(4-oxo-2-thioxothiazolidin-3-yl)pentanoic acid (**7e**). Pale yellow solid, 80% yield. Mp 74-76 °C. ¹H NMR (CDCl₃): δ 0.86 (t, *J* = 7 Hz, 3H, CH₃), 0.97-1.06 (m, 1H, CH_{2a}CH₃), 1.19 (d, *J* = 7 Hz, 3H, CHCH₃), 1.22-1.28 (m, 1H, CH_{2b}CH₃), 2.55 (m, 1H, CHCH₃), 4.02 (s, 2H, SCH₂), 5.31 (d, *J* = 9 Hz, 3H, NCH). ¹³C NMR (CDCl₃): δ 11.0, 17.5, 25.2, 33.4, 34.5, 61.9, 173.4, 173.6, 201.0. LR-ESI(-) *m/z* (%): 246 ([M-H]⁻, 23), 172 (100).

(S)-2-(4-oxo-2-thioxothiazolidin-3-yl)-3-phenylpropanoic acid (7f). Yellow solid, 89% yield. Mp 170-172 °C.

(lit.[4]170-173 °C). ¹H NMR (CDCl₃): δ 3.52 (d, J = 7 Hz, 2H, CH₂Ph), 3.77 (s, 2H, SCH₂), 5.85 (br s, 1H, NCH), 7.13-7.25 (m, 5H) 9.65 (s, 1H, CO₂H). LR-ESI(-) m/z (%): 280 ([M-H]⁻, 75).

Preparation of 2-Bromo-5-formylpyridine (8)

The compound was prepared following the literature procedure.[5] A suspension of 2-bromo-5-iodopyridine (3 g, 11 mmol) in dry Et₂O (100 mL) was cooled to -78 °C prior to the addition of *n*-BuLi (2.2 M, 5.3 mL, 1.1 equiv.). The reaction mixture was stirred for 1 h at -78 °C prior to the addition of dry DMF (1 mL). After stirring for 1 h, the reaction mixture was warmed to room temperature and quenched by the addition of dilute HCl (1 M, 20 mL). The organic layer was separated, and the aqueous layer was further extracted with Et₂O (2 x 20 mL). The ether layer was dried with MgSO₄, filtered, and the solvent removed in vacuo. Column chromatography on silica using a gradient (7-60% EtOAc, hexanes) yielded the pure product as a white solid (1.25 g, 64% yield). R_f (1:1 EtOAc/hexanes): 0.39. Mp 100-101°C. (lit.[6]100°C). ¹H NMR (CDCl₃): δ 7.69 (d, *J* = 8 Hz, 1H), 8.01 (dd, *J* = 8 Hz and 2 Hz, 1H), 8.83 (d, *J* = 2 Hz, 1H), 10.10 (s, 1H, CHO).

General Procedure for the Suzuki coupling of compound 8 to Boronic Acids 9a-d

To a solution of **8** (0.1 g, 0.54 mmol) in 1,4-dioxane (5 mL) was added the requisite boronic acid (1.1 equiv.) and aqueous K_3PO_4 (0.23g in 1 mL H₂O, 2 equiv.). The reaction mixture was degassed for 1 min with argon before adding catalytic $Pd_2(dba)_3$ (10 mg, 2 mol%) and PCy_3HBF_4 (8 mg, 4 mol%). The reaction mixture was stirred at 100 °C for 16 h in a sealed tube. The reaction mixture was then cooled to room temperature, and the solvent was removed. The residue was partitioned between EtOAc (20 mL) and water (20 mL), and the aqueous phase was further extracted with EtOAc (2 × 20 mL). Column chromatography on silica (EtOAc/hexanes, 0-50%) yielded the desired products as white or pale yellow solids.

2-(2,3-dimethoxyphenyl)-5-formylpyridine (**10a**). Pale yellow solid (95 % yield). Mp 84 °C.[7] ¹H NMR (CDCl₃): δ 3.72 (s, 3H, OCH₃), 3.93 (s, 3H, OCH₃), 7.04 (dd, J = 8 Hz and 2 Hz, 1H), 7.20 (t, J = 8 Hz, 1H), 7.44 (dd, J = 8 Hz and 2 Hz, 1H), 8.09 (d, J = 8 Hz, 1H), 8.20 (dd, J = 8 Hz and 2 Hz, 1H), 9.14 (d, J = 2 Hz, 1H), 10.14 (s, 1H). Anal (C₁₄H₁₃NO₃) C, H, N.

2-(3,4-methylenedioxyphenyl)-5-formylpyridine (**10b**). White solid (97 % yield). R_f (1: 3 EtOAc/hexanes): 0.33. Mp 133-138 °C. ¹H NMR (CDCl₃): δ 6.06 (s, 2H, CH₂), 6.94 (d, J = 9 Hz, 1H), 7.63 (m, 2H), 7.80 (d, J = 9 Hz, 1H), 8.18 (dd, J = 8 Hz and 2 Hz, 1H), 9.06 (d, J = 1 Hz, 1H), 10.11 (s, 1H, CHO). ¹³C NMR (CDCl₃): δ 101.6, 107.7, 108.6, 119.8, 122.2, 129.4, 132.3, 136.4, 148.5, 149.7, 152.4, 161.5, 190.3. FTIR (cm⁻¹, KBr): 757 w, 818 m, 846 w, 895 m, 931 m, 1037 s, 1107 m, 1240 s, 1302 m, 1369 s, 1404 m, 1476 s, 1558 m, 1591 s, 1696 s. LR-ESI(+) m/z (%): 228 ([M+H]⁺, 100). HR-ESI(+): Calcd 228.0655 for C₁₃H₁₀ NO₃ [M+H]⁺, found 228.0674. Anal (C₁₃H₉ NO₃) C, H, N.

2-(3,4-dimethoxyphenyl)-5-formylpyridine (**10c**). Yellow crystals (77 % yield). Mp 107-108 °C.[7] ¹H NMR (CDCl₃): δ 3.96 (s, 3H, OCH₃), 4.02 (s, 3H, OCH₃), 6.98 (d, J = 8 Hz, 1H), 7.63 (dd, J = 8 Hz and 2 Hz, 1H), 7.77 (d, J = 2 Hz, 1H), 7.86 (d, J = 8 Hz, 1H), 8.18 (dd, J = 8 Hz and 2 Hz, 1H), 9.07 (d, J = 2 Hz, 1H), 10.10 (s, 1H). Anal (C₁₄H₁₃NO₃) C, H, N.

2-(3-chloro-4-isopropoxyphenyl)-5-formylpyridine (**10d**). Colorless crystals (yield 93%). Mp 83 °C. ¹H NMR (CDCl₃): δ 1.43 (d, J = 6 Hz, 6H, 2 × CH₃), 4.68 (m, 1H, OCH), 7.05 (dd, J = 9 Hz, 1H), 7.82 (d, J = 8 Hz, 1H), 7.96 (dd, J = 8 Hz and 3 Hz, 1H), 8.15 (d, J = 3 Hz, 1H), 8.19 (dd, J = 8 Hz and 3 Hz, 1H), 9.07 (d, J = 2 Hz, 1H), 10.11 (s, 1H). ¹³C NMR (CDCl₃): δ 22.0, 72.0, 114.9, 119.7, 124.6, 126.9, 129.51, 129.57, 130.9, 136.5, 152.4, 155.5, 160.5, 190.3. HRMS (ESI⁺): Calcd 276.0786 for C₁₅H₁₅ClNO₂ [M+H]⁺, found 276.0785. Anal (C₁₅H₁₄ClNO₂) C, H, N.

General Procedure for the Knoevenagel Condensation of Aldehydes 10a-d with Rhodanines 7a-f

To a solution of the aldehydes **10a-d** (typically 100 mg, 0.30 mmol) in glacial acetic acid (1 to 5 mL) and sodium acetate (0.11 g, 4 equiv.) was added the desired rhodanine **7a-f** (2 equiv.). The mixture was refluxed for 3 h, then cooled to room temperature. The solvent was removed in vacuo, and aqueous HCl (1 M, 50 mL) was added and the resulting mixture was refluxed for a further 1 h. The reaction mixture was then cooled to room temperature, and the precipitate was collected via vacuum filtration. Recrystallization from hot 1 M HCl, or column chromatography on silica, gave the pure products **11a-x** (65-98% yields).

11a. Yellow powder. Mp 225-226 °C. ¹H NMR (DMSO- d_6): δ 3.70 (s, 3H, OCH₃), 3.87 (s, 3H, OCH₃), 4.73 (s, 2H, NCH₂), 7.20 (m, 2H), 7.38 (dd, J = 7 Hz and 3 Hz, 1H), 7.98 (s, 1H), 8.01 (d, J = 8 Hz, 1H), 8.09 (dd, J = 8 Hz and 2 Hz, 1H), 9.01 (d, J = 2 Hz, 1H). ¹³C NMR (DMSO- d_6): δ 45.1, 55.9, 60.6, 114.1, 122.0, 125.5, 124.2, 124.7, 127.3, 130.4, 132.4, 136.7, 147.0, 152.1, 152.9, 156.5, 166.2, 167.2, 192.8. FTIR (cm⁻¹, KBr): 536 w, 584 w, 786 w, 835 w, 847 w, 1125 m, 1222 s, 1265 m, 1342 s, 1387 m, 1405 m, 1470 m, 1491 m, 1595 m, 1780 s, 2833 w, 2944 w, 2987 w. Anal (C₁₉H₁₆N₂O₅S₂) C, H, N, S.

11b. Orange powder. Mp 208 °C. ¹H NMR (DMSO- d_6): δ 1.56 (d, J = 7 Hz, 3H, <u>CH</u>CH₃), 3.70 (s, 3H, OCH₃), 3.87 (s, 3H, OCH₃), 5.63 (q, J = 7 Hz, 1H, NCH), 7.20 (m, 2H), 7.38 (dd, J = 7 Hz and 3 Hz, 1H), 7.92 (s, 1H), 8.01 (d, J = 8 Hz, 1H), 8.06 (dd, J = 9 Hz and 2 Hz, 1H), 8.99 (d, J = 2 Hz, 1H). ¹³C NMR (DMSO- d_6): δ 13.4, 52.9, 55.9, 60.6, 114.1, 122.0, 123.1, 124.1, 124.6, 127.3, 130.1, 132.4, 136.6, 147.0, 152.0, 152.8, 156.4, 166.0, 169.5, 192.4. FTIR (cm⁻¹, KBr): 534 w, 745 m, 1003 m, 1026 m, 1114 m, 1235 s, 1270 s, 1470 m, 1591 m, 1724 s, 1911 br w, 2916 w. LR-ESI(-) m/z (%):429 ([M-H]⁻, 22), 386 (32), 271 (28), 126 (100). HR-ESI(-):Calcd. 429.0584 for C₂₀H₁₇N₂O₅S₂ [M-H]⁻, found 429.0566. Anal (C₂₀H₁₈N₂O₅S₂) C, H, N, calculated S, 14.90; found S, 14.13.

11c. Brown-yellow solid. Mp 222-224 °C. ¹H NMR (DMSO-*d*₆): δ 0.76 (d, *J* = 7 Hz, 3H, CH₃), 1.21 (d, *J* = 7 Hz, 3H, CH₃), 3.69 (s, 3H, OCH₃), 2.73 (m, 1H), 3.87 (s, 3H, OCH₃), 5.19 (d, *J* = 8 Hz, 1H), 7.19 (m, 2H), 7.38 (dd, *J* = 7 and 3 Hz, 1H), 7.96 (s, 1H), 8.00 (d, *J* = 8 Hz, 1H), 8.07 (dd, *J* = 8 and 2 Hz, 1H), 8.99 (d, *J* = 2 Hz, 1H). ¹³C NMR (DMSO-*d*₆): δ 18.9, 21.6, 27.1, 55.8, 60.5, 62.1, 114.1, 122.0, 122.3, 124.1, 124.6, 127.2, 130.9, 132.4, 136.7, 147.0, 152.0, 152.8, 156.5, 166.4, 168.5, 193.0. FTIR (KBr, cm⁻¹) 3423 (br), 2971, 1722, 1609, 1478, 1248, 1029, 833, 796, 751. HRMS (ESI⁻): Calcd 457.0897 for C₂₂H₂₁N₂O₅S₂ [M-H]⁻ Found: 457.0873. Anal (C₂₂H₂₂N₂O₅S₂) H, N, S calculated C, 57.82; found C, 56.20.

11d. Yellow solid. Mp 167-169 °C. ¹H NMR (CDCl₃): δ 0.95 (d, J = 6 Hz, 3H, CH₃), 1.00 (d, J = 6 Hz, 3H, CH₃), 1.57 (m, 1H), 2.16 (m, 1H), 2.30 (m, 1H), 3.71 (s, 3H, OCH₃), 3.93 (s, 3H, OCH₃), 5.78 (m, 1H, NCH), 7.03 (dd, J = 8 and 2 Hz, 1H), 7.21 (t, J = 8 Hz, 1H), 7.43 (dd, J = 8 and 2 Hz, 1H), 7.77 (s, 1H), 7.84 (dd, J = 8 and 3 Hz, 1H), 8.07 (d, J = 8 Hz, 1H), 8.88 (d, J = 3 Hz, 1H).¹³C NMR (CDCl₃): δ 22.2, 23.0, 25.3, 36.9, 56.0, 56.1, 61.2, 113.9, 122.6, 124.3, 124.6, 125.6, 127.8, 129.1, 132.2, 136.6, 147.5, 151.4, 153.1, 156.6, 167.1, 172.1, 192.2. FTIR (KBr, cm⁻¹) 3440 (br), 2963, 1729, 1590, 1263, 1206, 1028, 828, 795, 749. HRMS (ESI): Calcd 471.1054 for C₂₃H₂₃N₂O₅S₂ [M-H]⁻ Found: 471.1026. Anal. (C₂₃H₂₄N₂O₅S₂) C, H, N, S.

11e. Orange powder. Mp 129-130 °C. ¹H NMR (DMSO- d_6): δ 0.80 (t, J = 7 Hz, 3H, CH₂CH₃), 0.96 (m, 1H, CH_{2a}CH₃), 1.17 (d, J = 6 Hz, 3H, CHCH₃), 1.24 (m, 1H, CH_{2b}CH₃), 2.51 (m, 1H, CH₃CH), 3.69 (s, 3H, OCH₃), 3.87 (s, 3H, OCH₃), 5.21 (d, J = 9 Hz, 1H, NCHCO₂H), 7.20 (m, 2H), 7.39 (dd, J = 8 Hz and 1 Hz, 1H), 7.95 (s, 1H), 8.01 (m, 1H), 8.07 (dd, J = 9 Hz, 2H), 8.99 (d, J = 2 Hz, 1H). ¹³C NMR (CDCl₃): δ 11.2, 17.6, 25.3, 33.8, 56.0, 61.1, 62.0, 113.8, 122.6, 123.9, 124.6, 125.5, 127.7, 129.5, 132.4, 136.4, 147.5, 151.8, 153.1, 156.8, 167.3, 171.2, 192.2. FTIR (cm⁻¹, KBr): 548 w, 687 w, 738 w, 1001 m, 1036 m, 1094 w, 1126 m, 1236 br s, 1261 s, 1327 m, 1427 w, 1468 m, 1585 m, 1719 s, 2361 w, 2933 m, 2965 m. LR-ESI(-) m/z (%):471 ([M-H]⁻, 25), 428 (65), 357 (18), 168 (100). HR-ESI(-): Calcd. 471.1054 for C₂₃H₂₃N₂O₅S₂ [M-H]⁻, found 471.1038. Anal (C₂₃H₂₄N₂O₅S₂) C, H, N, S.

11f. Yellow solid. Mp 160-162 °C.[7] ¹H NMR (CDCl₃): δ 3.64 (d, J = 7 Hz, 2H), 3.70 (s, 3H, OCH₃), 3.92 (s, 3H, OCH₃), 6.00 (br s, 1H, NCH), 7.03 (d, J = 8 Hz, 1H), 7.20 (m, 6H), 7.40 (d, J = 8 Hz, 1H), 7.74 (s, 1H), 7.81 (d, J = 8 Hz, 1H), 8.04 (d, J = 8 Hz, 1H), 8.88 (s, 1H). Anal. (C₂₆H₂₂N₂O₅S₂) C, H, N, S.

11g. Pale orange powder. Mp 281-282 °C. ¹H NMR (DMSO- d_6): δ 4.76 (s, 2H, N<u>CH</u>₂CO₂H), 6.12 (s, 2H, OCH₂O), 7.06 (d, J = 8 Hz, 1H), 7.74 (d, J = 2 Hz, 1H), 7.77 (dd, J = 8 Hz and 2 Hz, 1H), 7.95 (s, 1H), 8.02 (dd, J = 9 Hz and 2 Hz, 1H), 8.09 (d, J = 9 Hz, 1H), 8.92 (d, J = 2 Hz, 1H). ¹³C NMR (DMSO- d_6): δ 45.1, 101.6, 106.8, 108.7, 119.9, 121.7, 122.8, 127.0, 130.5, 131.7, 137.5, 148.1, 149.1, 152.2, 156.6, 166.2, 167.2, 192.7. FTIR (cm⁻¹, KBr): 739 w, 807 w, 1037 m, 1105 m, 1196 s, 1251 m, 1270 m, 1328 s, 1393 m, 1439 m, 1472 s, 1584 s, 1712 s. LR-ESI(-) m/z (%): 399 ([M-H]⁻,73); 301 (91). HR-ESI(-): Calcd 399.0115 for C₁₈H₁₁N₂O₅S₂ [M-H]⁻, found: 399.0104. Anal (C₁₈H₁₂N₂O₅S₂) C, H, N, S.

11h. Orange-brown solid. Mp 251-252 °C. ¹H NMR (DMSO-*d*₆): δ 1.55 (d, *J* = 7 Hz, 3H, CH₃), 5.62 (q, *J* = 7 Hz, 1H, N<u>CH</u>CO₂H), 6.12 (s, 2H, OCH₂O), 7.07 (d, *J* = 8 Hz, 1H), 7.74 (d, *J* = 2 Hz, 1H), 7.77 (dd, *J* = 8 Hz and 2 Hz, 1H), 7.89 (s, 1H), 8.01 (dd, *J* = 9 Hz and 1 Hz, 1H), 8.10 (d, *J* = 8 Hz, 1H), 8.91 (d, *J* = 2 Hz, 1H).¹³C NMR (DMSO-*d*₆): δ 13.4, 52.9, 101.6, 106.8, 108.7, 119.9, 121.7, 122.4, 127.0, 130.2, 131.7, 137.4, 148.1, 149.1, 152.1, 156.5, 166.0, 169.5, 192.3. Anal (C₁₉H₁₄N₂O₅S₂) Calcd C, 55.80; H, 4.21; N, 6.17; S, 14.90. Found C, 53.04; H, 4.25; N, 6.01; S, 15.16. FTIR (cm⁻¹, KBr): 467 w, 549 w, 597 w, 636 w, 686 w, 764 w, 808 m, 846 w, 894 m, 917 m, 935 m, 1001 w, 1036 m, 1053 m, 1108 m, 1144 w, 1246 s, 1257 s, 1305 m, 1348 s, 1391 m, 1442 m, 1476 s, 1501 m, 1591 m, 1608 m, 1700 s, 1740 m.LR-ESI(-) *m*/*z* (%): 413 ([M-H]⁻, 8); 249 (13); 155 (32); 123 (100); 69 (57). HRMS: Calcd 413.0271 for C₁₉H₁₃N₂O₅S₂ [M-H]⁻; found 413.0286. Anal (C₁₉H₁₄N₂O₅S₂) C, H, N, S.

11i. Yellow solid. Mp 248 °C. ¹H NMR (DMSO- d_6): δ 0.76 (d, J = 7 Hz, 3H, CH₃), 1.21 (d, J = 7 Hz, 3H, CH₃), 2.74 (m, 1H, <u>CH</u>(CH₃)₂), 5.18 (d, J = 7 Hz, 1H, N<u>CH</u>CO₂H) 6.12 (s, 2H, OCH₂O), 7.07 (d, J = 8 Hz, 1H), 7.75 (d, J = 2 Hz, 1H), 7.78 (dd, J = 8 Hz and 2 Hz, 1H), 7.94 (s, 1H), 8.03 (dd, J = 9 Hz and 2 Hz, 1H), 8.11 (d, J = 9 Hz, 1H), 8.92 (d, J = 2 Hz, 1H). ¹³C NMR (DMSO- d_6): δ 18.9, 21.6, 27.1, 62.1, 101.6, 106.7, 108.6, 119.8, 121.5, 121.7, 126.9, 131.0, 131.6, 137.5, 148.1, 149.1, 152.2, 156.6, 166.4, 168.5, 192.9. FTIR (cm⁻¹, KBr): 547 w, 808 w, 1037 m, 1201 m, 1237 s, 1330 m, 1347 m, 1475 s, 1591 m, 1607 m, 1702 s, 1734 s, 2973 w. LR-ESI (-) m/z (%): 441

([M-H]⁻, 50); 398 (100); 342 (30). HRMS: Calcd 441.0584 for $C_{21}H_{17}N_2O_5S_2$ [M-H]⁻; found 441.0568. Anal ($C_{21}H_{18}N_2O_5S_2$) C, H, N, S.

11j. Yellow solid. Mp 233 °C. ¹H NMR (DMSO- d_6): δ 0.87 (d, J = 7 Hz, 3H, CH₃), 0.92 (d, J = 7 Hz, 3H, CH₃), 1.48 (m, 1H, <u>CH_{2a}</u>CH), 2.03 (m, 1H, <u>CH</u>(CH₃)₂), 2.21 (m, 1H, <u>CH_{2b}</u>CH), 5.60 (br s, 1H, N<u>CH</u>CO₂H), 6.12 (s, 2H, OCH₂O), 7.07 (d, J = 8 Hz, 1H), 7.74 (d, J = 2 Hz, 1H), 7.76 (dd, J = 8 Hz and 2 Hz, 1H), 7.91 (s, 1H), 8.01 (dd, J = 9 Hz and 2 Hz, 1H), 8.09 (d, J = 9 Hz, 1H), 8.91 (d, J = 2 Hz, 1H). ¹³C NMR (DMSO- d_6): δ 21.9, 22.8, 24.8, 36.3, 55.9, 101.6, 106.7, 108.6, 119.9, 121.7, 121.9, 127.0, 130.6, 131.6, 137.5, 148.1, 149.1, 152.2, 156. 6, 166.4, 169.3, 193.0. FTIR (cm⁻¹, KBr): 747 w, 813 m, 1036 s, 1226 s, 1249 s, 1268 s, 1337 s, 1390 m, 1475 s, 1593 m, 1608 m, 1699 s, 1734 br m, 2364 w, 2905 w. LR-ESI(-) m/z (%): 455 ([M-H]⁻, 22); 339 (12); 325 (52); 297 (14); 243 (15); 192 (43). HR-ESI(-): Calcd 455.0741 for C₂₂H₁₉N₂O₅S₂ [M-H]⁻, found 455.0724. Anal (C₂₂H₂₀N₂O₅S₂) C, H, N, S.

11k. Yellow-orange flakes. Mp 221-222 °C. ¹H NMR (DMSO-*d*₆): δ 0.80 (t, *J* = 7 Hz, 3H, CH₂<u>CH₃</u>), 0.95 (m, 1H, <u>CH_{2a}</u>CH₃), 1.16 (d, *J* = 7 Hz, 3H, <u>CH</u>CH₃), 1.23 (m, 1H, <u>CH_{2b}</u>CH₃), 2.51 (m, 1H, CH₃<u>CH</u>), 5.22 (d, *J* = 9 Hz, 1H, N<u>CH</u>CO₂H), 6.12 (s, 2H, OCH₂O), 7.07 (d, *J* = 8 Hz, 1H), 7.74 (d, *J* = 2 Hz, 1H), 7.78 (dd, *J* = 8 Hz and 2 Hz, 1H), 7.93 (s, 1H), 8.02 (dd, *J* = 9 Hz and 2 Hz, 1H), 8.11 (d, *J* = 8 Hz, 1H), 8.92 (d, *J* = 2 Hz, 1H). ¹³C NMR (DMSO-*d*₆): δ 10.9, 17.5, 24.9, 33.0, 61.6, 79.1, 101.6, 106.8, 108.7, 119.9, 121.5, 127.0, 131.0, 131.7, 137.6, 148.2, 149.2, 152.2, 156.6, 166.5, 168.6, 193.0. FTIR (cm⁻¹, KBr): 547 w, 583 w, 660 w, 691 w, 740 w, 808 m, 845 w, 893 w, 936 w, 958 w, 1037 m, 1102 m, 1132 m, 1201 m, 1237 s, 1305 m, 1330 m, 1347 m, 1391 m, 1440 m, 1475 s, 1501 m, 1555 w, 1591 m, 1607 m, 1702 s, 1734 m, 2363 m, 2929 m, 2973 m. LR-ESI(-) *m/z* (%): 445 ([M-H]⁻, 33); 412 (78); 341 (22); 256 (25); 168 (75). HR-ESI(-): Calcd 455.0741 for C₂₂H₁₉N₂O₅S₂ [M-H]⁻, found: 455.0739. Anal (C₂₂H₂₀N₂O₅S₂) C, H, N, S.

111. Yellow powder. Mp 229 °C. ¹H NMR (DMSO-*d*₆): δ 3.52 (d, *J* = 5 Hz, 2H, CH₂Ph), 5.90 (br s, 1H, N<u>CH</u>CO₂H), 6.11 (s, 2H, OCH₂O), 7.05 (d, *J* = 8 Hz, 1H), 7.13-7.23 (m, 5H), 7.73 (d, *J* = 2 Hz, 1H), 7.76 (dd, *J* = 8 Hz and 2 Hz, 1H), 7.86 (s, 1H), 7.95 (dd, *J* = 9 Hz and 2 Hz, 1H), 8.06 (d, *J* = 9 Hz, 1H), 8.88 (d, *J* = 2 Hz, 1H). ¹³C NMR (DMSO-*d*₆): δ 33.0, 58.2, 101.6, 106.7, 108.6, 119.8, 121.5, 121.7, 126.7, 126.8, 128.3, 128.9, 130.5, 131.6, 136.4, 137.5, 148.1, 149.1, 152.2, 156.6, 166.3, 168.6, 192.2. FTIR (cm⁻¹, KBr): 839 w, 1035 m, 1170 m, 1235 s, 1266 s, 1338 s, 1475 s, 1592 m, 1609 m, 1706 s, 1732 br m, 2890 w. LR-ESI(-) *m/z* (%): 489 ([M-H]⁻, 100); 445 ([M-C₃H₈]⁻, 85); 354 (28); 311 (43); 243 (32); 219 (16). HR-ESI(-): Calcd 489.0584 for C₂₅H₁₇N₂O₅S₂ [M-H]⁻, found 489.0593. Anal (C₂₅H₁₈N₂O₅S₂) C, H, N, S.

11m. Orange solid. Mp 177-178 °C (dec). ¹H NMR (DMSO- d_6): δ 3.84 (s, 3H, OCH₃), 3.87 (s, 3H, OCH₃), 4.77 (s, 2H, NCH₂), 7.10 (d, J = 8 Hz, 1H), 7.79 (m, 2H), 7.97 (s, 1H), 8.04 (dd, J = 8 Hz and 2 Hz, 1H), 8.15 (d, J = 8 Hz, 1H), 8.95 (d, J = 2 Hz, 1H). ¹³C NMR (DMSO- d_6): δ 45.1, 55.5, 55.6, 110.0, 111.8, 120.2, 120.3, 122.9, 127.0, 129.2, 130.3, 138.0, 149.0, 151.0, 151.5, 156.3, 166.2, 167.2, 192.6. FTIR (KBr, cm⁻¹) 3445, 2939, 2843, 1722, 1586, 1516, 1332, 1201, 1058, 842, 734, 609. HRMS (ESI'): Calcd 415.0428 for C₁₉H₁₅N₂O₅S₂ [M-H]⁻ Found: 415.0418. Anal (C₁₉H₁₆N₂O₅S₂) Calcd C, 54.79; H, 3.87; N, 6.73; S, 15.40. Found C, 45.84; H, 3.72; N, 5.61; S, 12.99.

11n. Brown-yellow solid. Mp 243-244 °C. ¹H NMR (DMSO-*d*₆): δ 1.56 (d, *J* = 7 Hz, 3H, CH₃), 3.83 (s, 3H, OCH₃), 3.86 (s, 3H, OCH₃), 5.63 (q, *J* = 7 Hz, 1H, NCH), 7.10 (d, *J* = 8 Hz, 1H), 7.77 (m 2H), 7.90 (s, 1H), 8.00 (dd, *J* = 8 and 2 Hz, 1H), 8.14 (d, *J* = 8 Hz, 1H), 8.92 (d, *J* = 2 Hz, 1H). ¹³C NMR (DMSO-*d*₆): δ 13.4, 52.8, 55.5, 55.6, 109.9, 111.7, 119.8, 120.0, 122.2, 126.8, 129.9, 130.4, 137.4, 149.0, 150.8, 152.8, 156.9, 166.0, 169.5, 192.3. FTIR (KBr, cm⁻¹) 3424 (br), 2938, 2361, 1721, 1584, 1287, 1238, 814, 731, 669. HRMS (ESI⁻): Calcd 429.0584 for C₂₀H₁₇N₂O₅S₂ [M-H]⁻ Found: 429.0573. Anal (C₂₀H₁₈N₂O₅S₂) Calcd C, 55.80; H, 4.21; N, 6.51; S, 14.90. Found C, 53.04; H, 4.25; N, 6.01; S, 15.16.

110. Brown-yellow solid. Mp 182-184 °C. ¹H NMR (DMSO-*d*₆): δ 0.76 (d, *J* = 7 Hz, 3H, CH₃), 1.21 (d, *J* = 6 Hz, 3H, CH₃), 2.75 (m, 1H, CH), 3.84 (s, 3H, OCH₃), 3.87 (s, 3H, OCH₃), 5.19 (d, *J* = 9 Hz, 1H, NCH), 7.11 (d, *J* = 8 Hz, 1H), 7.78 (m, 2H), 7.95 (s, 1H), 8.04 (dd, *J* = 9 and 2 Hz, 1H), 8.16 (d, *J* = 9 Hz, 1H), 8.94 (d, *J* = 2 Hz, 1H). ¹³C NMR (DMSO-*d*₆): δ 19.0, 21.7, 27.2, 55.5, 55.6, 62.2, 109.9, 111.8, 119.8, 120.1, 121.4, 126.8, 130.0, 131.2, 137.6, 149.0, 150.9, 152.3, 157.0, 166.5, 168.6, 193.0. FTIR (KBr, cm⁻¹) 3550 (br), 2965, 2363, 1726, 1585, 1244, 1026, 836, 737. HRMS (ESI⁻): Calcd 457.0897 for C₂₂H₂₁N₂O₅S₂ [M-H]⁻ Found: 457.0882. Anal (C₂₂H₂₂N₂O₅S₂) Calcd C, 57.62; H, 4.84; N, 6.11;S, 13.99. Found C, 52.40; H, 5.25; N, 5.04; S, 17.93.

11p. Yellow solid. Mp 162-164 °C. ¹H NMR (CDCl₃): δ 0.95 (d, J = 7 Hz, 3H, CH₃), 1.00 (d, J = 7 Hz, CH₃), 1.56 (m, 1H), 2.15 (m, 1H), 2.30 (m, 1H), 3.95 (s, 3H, OCH₃), 4.00 (s, 3H, OCH₃), 5.80 (m, 1H, NCH), 6.97 (d, J = 9 Hz, 1H), 7.56 (dd, J = 9 Hz and 1 Hz, 1H), 7.69 (br s, 1H), 7.73 (s, 1H), 7.81 (br s, 2H), 8.82 (s, 1H). ¹³C NMR (CDCl₃): δ 22.2, 23.0, 25.2, 36.9, 56.00, 56.03, 56.1, 110.0, 111.1, 120.3, 120.4, 123.3, 127.2, 129.4, 130.4, 137.1, 149.5, 151.1, 152.0, 157.8, 167.0, 172.5, 192.1. FTIR (KBr, cm⁻¹) 3450 (br), 2959, 1722, 1584, 1275, 1208, 1025,

834, 770, 738. HRMS (ESI): Calcd 471.1054 for $C_{23}H_{23}N_2O_5S_2$ [M-H]⁻ Found: 471.1033. Anal ($C_{23}H_{24}N_2O_5S_2$) C, H, N, S.

11q. Orange solid. Mp 157-158 °C. ¹H NMR (DMSO- d_6): δ 0.80 (t, J = 7 Hz, 3H, CH₃), 0.95 (m, 1H), 1.16 (d, J = 6 Hz, 3H, CH₃), 1.24 (m, 1H), 2.54 (m, 1H), 3.87 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 5.24 (d, J = 9 Hz, 1H, NCH), 7.10 (d, J = 8 Hz, 1H), 7.79 (m, 2H), 7.94 (s, 1H), 8.03 (dd, J = 9 and 2 Hz, 1H), 8.15 (d, J = 9 Hz, 1H), 8.93 (d, J = 2 Hz, 1H), 13.29 (br s, 1H, CO₂H). ¹³C NMR (DMSO- d_6): δ 10.9, 17.5, 24.8, 33.0, 55.5, 55.6, 61.6, 109.9, 111.7, 119.8, 120.1, 121.3, 126.8, 129.9, 131.2, 137.5, 149.0, 150.8, 152.3, 157.0, 166.5, 168.6, 193.0. FTIR (KBr, cm⁻¹) 3480 (br), 2963, 2361, 1716, 1583, 1235, 1026, 836, 770, 736. HRMS (ESI⁻): Calcd 471.1054 for C₂₃H₂₃N₂O₅S₂ [M-H]⁻ Found: 471.1040. Anal. (C₂₃H₂₄N₂O₅S₂) C, H, N, S.

11r. Orange solid. Mp 202-203 °C.[7] ¹H NMR (DMSO- d_6): δ 3.53 (d, J= 4 Hz, 2H, PhCH₂), 3.83 (s, 3H, CH₃), 3.86 (s, 3H, CH₃), 5.86 (br s, 1H, NCH), 7.09 (d, J= 8 Hz, 1H), 7.15-7.23 (m, 5H), 7.77 (m, 2H), 7.86 (s, 1H), 7.98 (dd, J= 2 and 9 Hz, 1H), 8.13 (d, J= 9 Hz, 1H), 8.90 (d, J= 2 Hz, 1H). Anal (C₂₆H₂₃ClN₂O₅S₂) C, H, N, S.

11s. Brown-yellow solid. Mp 236-237 °C. ¹H NMR (DMSO-*d*₆): δ 1.34 (d, *J* = 6 Hz, 6H, 2 × CH₃), 4.76 (s, 2H, NCH₂), 4.81 (m, 1H), 7.32 (d, *J* = 9 Hz, 1H), 7.97 (s, 1H), 8.06 (dd, *J* = 9 and 2 Hz, 1H), 8.15 (dd, *J* = 9 and 2 Hz, 1H), 8.18 (d, *J* = 9 Hz, 1H), 8.26 (d, *J* = 2 Hz, 1H), 8.96 (d, *J* = 2 Hz, 1H). ¹³C NMR (DMSO-*d*₆): δ 21.8, 45.1, 71.3, 115.2, 119.9, 122.9, 123.0, 127.0, 127.3, 128.5, 130.5, 137.6, 152.3, 154.5, 155.5, 166.2, 167.3, 192.6. FTIR (KBr, cm⁻¹) 2980, 1708, 1582, 1321, 1199, 1109, 951, 812, 743, 617. HRMS (ESI⁻): Calcd 447.0246 for C₂₀H₁₆ClN₂O₄S₂ [M-H]⁻ Found: 447.0231. Anal (C₂₀H₁₇ClN₂O₄S₂) Calcd C, 53.51; H, 3.82; N, 6.24; S, 14.28. Found C, 52.14; H, 3.85; N, 5.82; S, 13.30.

11t. Yellow solid. Mp 182-183 °C. ¹H NMR (DMSO- d_6): $\delta 1.34$ (d, J = 7 Hz, 6H, 2 × CH₃), 1.56 (d, J = 7 Hz, 3H, CH₃), 4.81 (m, 1H), 5.63 (q, J = 7 Hz, 1H, NCH), 7.32 (d, J = 9 Hz, 1H), 7.91 (s, 1H), 8.04 (dd, J = 8 and 2 Hz, 1H), 8.13 (m, 2H), 8.26 (d, J = 2 Hz, 1H), 8.94 (d, J = 2 Hz, 1H). ¹³C NMR (CDCl₃): $\delta 13.7$, 22.0, 53.1, 72.1, 115.1, 120.2, 123.9, 124.7, 126.8, 127.5, 129.3, 129.4, 130.6, 137.4, 151.9, 155.4, 156.7, 166.6, 172.6, 191.4. FTIR (KBr, cm⁻¹) 2978, 2361, 1711, 1582, 1475, 1247, 1109, 950, 811, 736. HRMS (ESI⁻): Calcd 461.0402 for C₂₁H₁₈CIN₂O₄S₂ [M-H]⁻ Found: 461.0377. Anal (C₂₁H₁₉CIN₂O₄S₂) C, H, N, S.

11u. Yellow solid. Mp 223-225 °C. ¹H NMR (CDCl₃): δ 0.85 (d, J = 7 Hz, 3H, CH₃), 1.29 (d, J = 6 Hz, 3H, CH₃), 1.41 (s, 3H, CH₃), 1.43 (s, 3H, CH₃), 2.91 (m, 1H), 4.66 (m, 1H), 5.38 (d, J = 9 Hz, 1H, NCH), 7.04 (d, J = 9 Hz, 1H), 7.73 (s, 1H), 7.78 (d, J = 8 Hz, 1H), 7.83 (dd, J = 9 and 2 Hz, 1H), 7.90 (dd, J = 8 and 2 Hz, 1H), 8.06 (d, J = 2 Hz, 1H), 8.82 (d, J = 2 Hz, 1H). ¹³C NMR (CDCl₃): δ 19.1, 21.7, 22.0, 27.7, 62.4, 72.1, 115.1, 120.2, 123.6, 124.7, 126.8, 127.5, 129.35, 129.37, 130.6, 137.4, 152.0, 155.4, 156.7, 167.2, 171.9, 192.1. HRMS (ESI⁻): Calcd: 489.0715 for C₂₃H₂₂ClN₂O₄S₂ [M-H]⁻ Found: 489.0738. Anal (C₂₃H₂₃ClN₂O₄S₂) C, H, N, S.

11v. Yellow solid. Mp 213-216 °C. ¹H NMR (CDCl₃): δ 0.94 (d, J = 7 Hz, 3H, CH₃). 0.99 (d, J = 7 Hz, 3H, CH₃), 1.41 (s, 3H, CH₃), 1.43 (s, 3H, CH₃), 1.56 (m, 1H), 2.15 (m, 1H), 2.31 (m, 1H), 4.66 (m, 1H, OCH), 5.80 (m, 1H, NCH), 7.04 (d, J = 9 Hz, 1H), 7.80 (d, J = 8 Hz, 1H), 7.72 (s, 1H), 7.82 (dd, J = 8 and 2 Hz, 1H), 7.91 (dd, J = 9 and 2 Hz, 1H), 8.08 (d, J = 2 Hz, 1H), 8.81 (d, J = 2.2 Hz, 1H). ¹³C NMR (CDCl₃): δ 22.0, 22.2, 23.0, 25.2, 36.9, 56.0, 72.1, 115.1, 120.1, 123.7, 124.7, 126.7, 127.5, 129.2, 129.3, 130.6, 137.4, 151.9, 155.4, 156.7, 167.0, 172.8, 192.0. FTIR (KBr, cm⁻¹) 2962, 1720, 1584, 1475, 1281, 1208, 1107, 952, 814, 736. HRMS (ESI⁻): Calcd 503.0872 for C₂₄H₂₄ClN₂O₄S₂ [M-H]⁻ Found: 503.0897. Anal. (C₂₄H₂₅ClN₂O₄S₂)C, H, N, S.

11w. Yellow solid. Mp 157-159 °C. ¹H NMR (CDCl₃): δ 0.88 (t, *J* = 7Hz, 3H), 1.07 (m, 1H), 1.25 (d, *J* = 7 Hz, 3H, CH₃), 1.31 (m, 1H), 1.43 (d, *J* = 6 Hz, 6H, 2 × CH₃), 2.66 (m, 1H), 4.67 (m, 1H), 5.44 (d, *J* = 9 Hz, 1H, NCH), 7.05 (d, *J* = 9 Hz, 1H), 7.74 (s, 1H), 7.79 (d, *J* = 9 Hz, 1H), 7.83 (dd, *J* = 9 and 2 Hz, 1H), 7.92 (dd, *J* = 9 and 2 Hz, 1H), 8.07 (d, *J* = 2 Hz, 1H), 8.81 (d, *J* = 2 Hz, 1H). ¹³C NMR (CDCl₃): δ 11.1, 17.6, 22.0, 25.3, 33.7, 61.9, 72.1, 115.1, 120.1, 123.5, 124.7, 126.7, 127.5, 129.3, 129.4, 130.6, 137.3, 152.0, 155.4, 156.7, 167.2, 171.8, 192.1. FTIR (KBr, cm⁻¹) 2973, 2361, 1721, 1584, 1475, 1236, 1106, 951, 813, 734, 688. HRMS (ESI⁻): Calcd 503.0872 for C₂₄H₂₄ClN₂O₄S₂ [M-H]⁻ Found: 503.0853. Anal (C₂₄H₂₅ClN₂O₄S₂) C, H, N, S.

11x. Yellow solid. Mp 181-183 °C. ¹H NMR (DMSO- d_6): δ 1.33 (d, J = 6 Hz, 6H, 2 × CH₃), 3.53 (d, J = 5 Hz, 2H), 4.78 (m, 1H, OCH), 5.90 (br s, 1H, NCH), 7.18 (m, 5H), 7.29 (d, J = 9 Hz, 1H), 7.86 (s, 1H), 7.97 (dd, J = 9 and 2 Hz, 1H), 8.10 (dd, J = 9 and 3 Hz, 1H), 8.11 (d, J = 9 Hz, 1H), 8.23 (d, J = 2 Hz, 1H), 8.89 (d, J = 3 Hz, 1H). ¹³C NMR (DMSO- d_6): δ 21.7, 33.0, 58.2, 71.2, 115.1, 119.8, 121.7, 122.8, 126.7, 126.9, 127.1, 128.2, 128.4, 128.9, 130.4, 136.5, 137.6, 152.3, 154.5, 155.4, 166.3, 168.6, 192.2. FTIR (KBr, cm⁻¹) 2978, 1720, 1583, 1475, 1279, 1107, 950, 815, 736. HRMS (ESI'): Calcd 537.0715 for C₂₇H₂₂ClN₂O₄S₂ [M-H]⁻ Found: 537.0695. Anal. (C₂₇H₂₃ClN₂O₄S₂) C, H, N, S.

Molecular Docking

The X-ray crystal structures of Bcl-XL and Mcl-1 with the Bim-BH3 peptide were downloaded from the PDB database (PDB IDs 3DFL and 2PQK). Sequence alignents and percent identity were confirmed using the matchmaker scripts in UCSF Chimera version 1.3.[8] The 3D structures of the inhibitors were generated using ChemBio 3D Ultra followed by MMFF energy minimization. AutoDock 4.0.1 and ADT were used for generating the docking models.[9, 10] Grid maps covering the entire BH3 binding site of both proteins were used in AutoDock calculations using the standard grid spacing of 0.375 Å. The GA-LS algorithm was adopted using the default setting except for the maximum number of energy evaluations which was set to 1,000,000. For each docking job, 100 hybrid GA-LS runs were employed. A total of 100 possible binding conformations were generated and grouped into clusters using a 1.0 Å root-mean-square tolerance. The lowest energy clusters were selected to create the docking poses, and the results were analyzed using ADT.

Biology. Fluorescence Polarization Assay

The Flu-Bak-BH3 peptide was purchased from Mimotopes (Clayton, Victoria, Australia). The peptide was prepared as 1 mM stock solution in DMSO, and stock solutions of the test compounds (4 mM in DMSO) were used for serial dilutions (250 μ M to 0.65 μ M final concentrations). The assay was carried out in a total volume of 100 μ L/well containing 3 μ g glutathione *S*-transferase (GST)-hBcl_{XL} Δ C19 or 3 μ g glutathione *S*-transferase (GST)-hMcl-1 Δ C20 and 60 nM labeled peptide in buffer (50 mM Tris, pH 8, 150 mM NaCl and 0.1% bovine serum albumin). In each well was added 10 μ L of the test compounds, and the reaction mixture was incubated at room temperature for 1 h. Each compound was tested in quadruplicate. The fluorescence polarization values were determined using Tecan GeniosPro plate reader using the excitation/emission wavelengths 485/535 nm. The data was analyzed using GraphPad Prism 5 and the results were fitted to a dose-response curve to obtain the IC₅₀ values and standard deviations. The K_i values were calculated using an online calculator[11] using the following parameters: [L]= 0.06 μ M for Flu-Bak, [Bcl-XL]=1.1 μ M, and [Mcl-1]= 1.6 μ M. The dissociation constants employed were K_d (Bcl-XL)= 0.14 μ M and K_d (Mcl-1)= 1.1 μ M.

References

- 1. H. Korner. Chem. Ber. 41, 1901-1905. (1908).
- 2. B. Holmberg. Compt. rend. trav. lab. Carlsberg, Ser. chim. 22, 211-218. (1938).
- 3. I. I. Kopiichuk. Farm. Zh. (Kiev). 21, 7-10 (1966).
- 4. F. Zuber, E. Sorkin. Helv. Chim. Acta 35, 1744-1747 (1952).
- 5. M. v. d. B. van den Heuvel, Tieme A.; Kellogg, Richard M.; Choma, Christin T.; Feringa, Ben
- L. J. Org. Chem. 69, 250-262 (2004).
- 6. P. M. Windscheif, F. Voegtle. Synthesis, 87-92 (1994).
- 7. P. H. Bernardo, T. Sivaraman, K.-F. Wan, J. Xu, J. Krishnamoorthy, C. M. Song, L. Tian, J. S. F. Chin, D. S. W. Lim, H. Y. K. Mok, V. C. Yu, J. C. Tong, C. L. L. Chai. *Journal of Medicinal Chemistry* **53**, 2314-2318 (2010).
- 8. E. F. Pettersen, T. D. Goddard, C. C. Huang, G. S. Couch, D. M. Greenblatt, E. C. Meng, T. E. Ferrin. *J. Comput. Chem.* **25**, 1605-1612 (2004).
- 9. M. F. Sanner. J. Mol. Graphics Mod. 17, 57-61 (1999).
- 10. M. F. Sanner, J.-C. Spehner, A. J. Olson. Biopolymers 38, 305-320 (1996).
- 11. <u>http://botdb.abcc.ncifcrf.gov/toxin/kiConverter.jsp</u>.